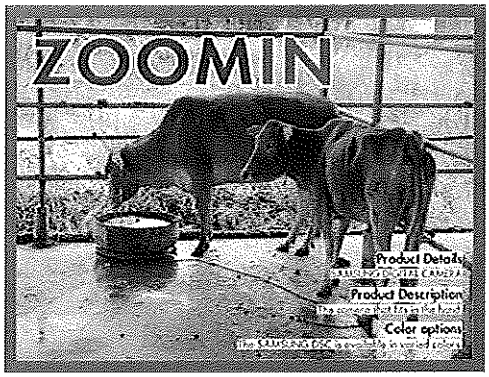
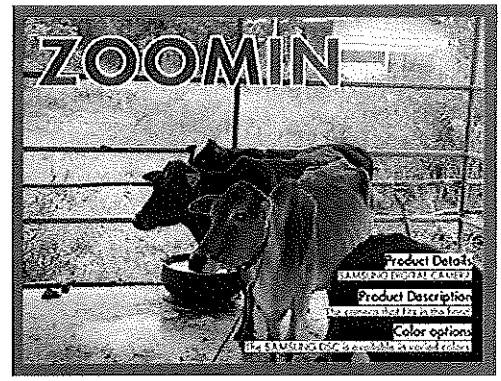
ภาคผนวก



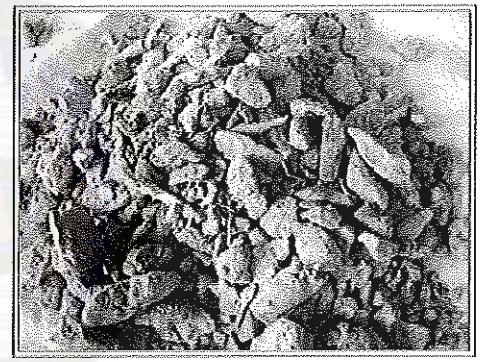
กลุ่ม ควบคุม



กลุ่ม ทดลอง

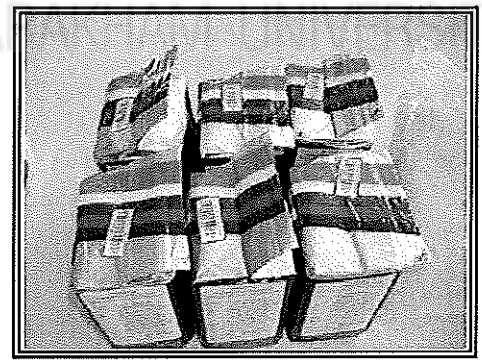
ภาพที่ 1 แสดงสัตว์ทดลองจำนวน 4 ตัว

ระยอง 7



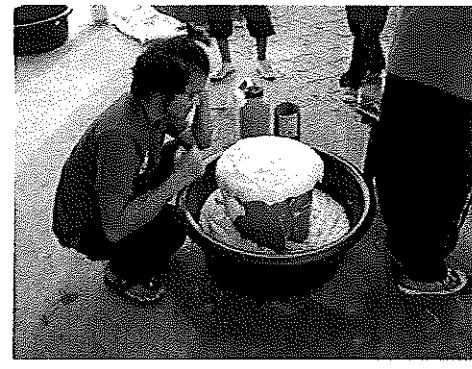
มันเส้น

น้ำตาลทราย



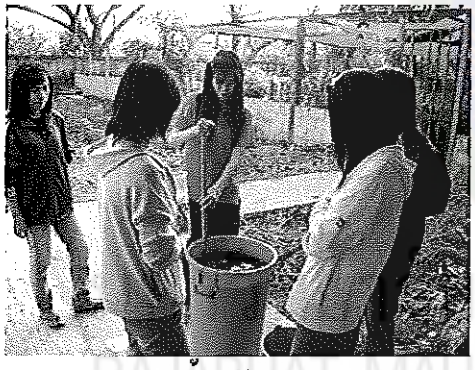
เชื้อยีสต์ baker yeast (*Saccharomyces cerevisiae*)

ภาพที่ 2 แสดงวัตถุดิบการทำมันเส้นหมักยีสต์-มาเลท

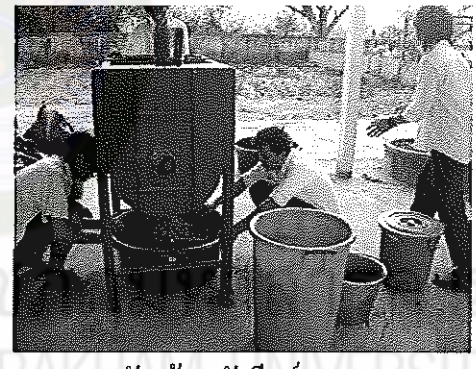


กระบวนการกระตุ้นเชื้อยีสต์

ภาพที่ 3 แสดงขั้นตอนการเตรียม ยีสต์หมัก

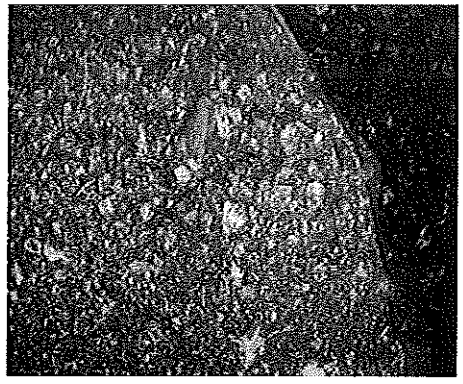


ผสมน้ำหมัก



ผสมมันเส้นหมักยีสต์-มาเลท

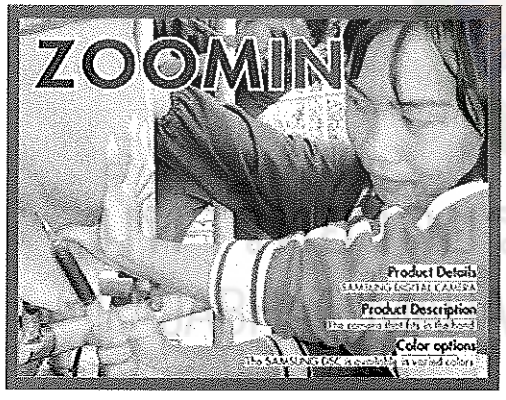
ภาพที่ 4 แสดงขั้นตอนการทำน้ำหมักยีสต์



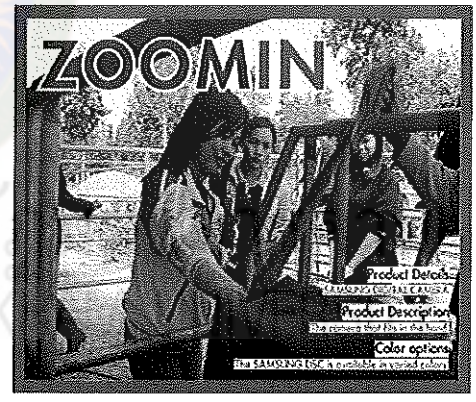
กระบวนการผสมมันเส้นหมักยีสต์-มาเลท

การตากแห้ง

ภาพที่ 5 แสดงขั้นตอนการผสมมันเส้นหมักยีสต์-มาเลท

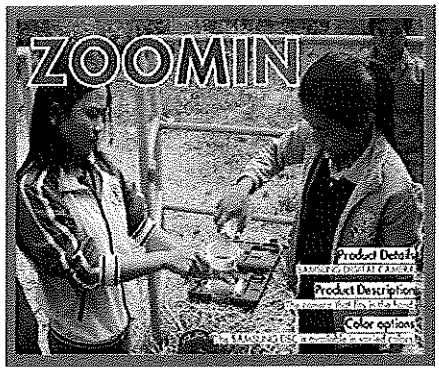


เก็บตัวอย่างเลือด

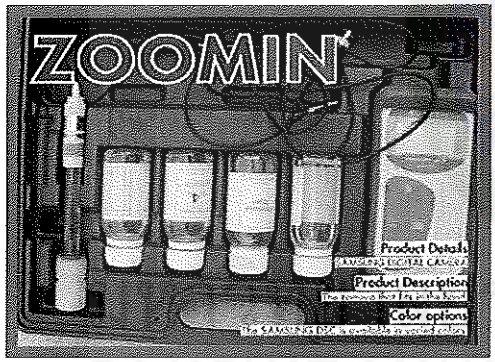


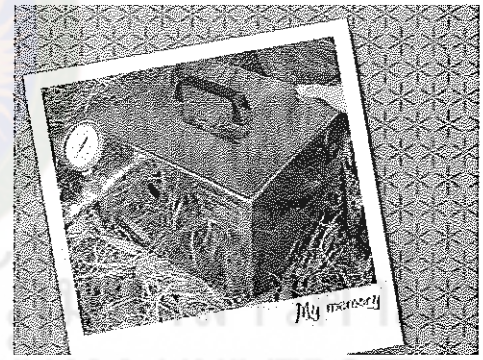
เก็บตัวอย่างอุจาระ

ภาพที่ 6 การเก็บตัวอย่างเพื่อการวิเคราะห์และประเมินผล



การเก็บของเหลวในกระเพาะหมัก





เครื่องวัดค่า pH

เครื่อง Suction pump

ภาพที่ 7 แสดงขั้นตอนการเก็บตัวอย่างของเหลวในกระเพาะหมัก (Rumen fluid)

ภาคผนวก

Pakistan Journal of Nutrition 8 (5): 597-600, 2009
ISSN 1680-5194

Supplementation of Yeast Fermented Cassava Chip (YFCC) as a Replacement Concentrate and Ruzi Grass on Rumen Ecology in Native Cattle

Sittisak Khampa[1], Pala Chaowarat[1], Rungson Singhalert[2] and Metha Wanapat[3]
[1]Faculty of Agricultural Technology, [2]Faculty of Humanities and Social Sciences,
Rajabhat Mahasarakham University, P.O. Box 44000, Thailand,
[3]Tropical Feed Resources Research and Development Center (TROFREC), Department of Animal Science,
Faculty of Agriculture, Khon Kaen University, P.O. Box 40002, Thailand

Abstract: Ten, one year old of native cattle with initial body weight of 150 ± 10 kg were randomly divided into two groups and received concentrate at 14% CP (T1) and Yeast Fermented Cassava Chip (YFCC) (T2). The cows were offered the treatment concentrate at 1% BW and ruzi grass was fed *ad libitum*. Means were compared using T-test. All animals were kept in individual pens and received free access to water. The results have revealed that replacement of YFCC on feed intake was non-significantly different, while Average Daily Gain (ADG) was higher ($p<0.05$) in native cattle fed YFCC (T2) treatments than received concentrate at 14% CP (T1) (259 and 205 g/d). In addition, the ruminal pH, ammonia-nitrogen and blood urea nitrogen concentration were significantly different ($p<0.05$). Supplementation of YFCC (T2) could improve population of bacteria and fungal zoospore, but decreased populations of *Holotrich* and *Entodiniomorph* protozoa in rumen ($p<0.05$). The results indicate that supplementation of Yeast Fermented Cassava Chip (YFCC) as a replacement concentrate at 14% CP could improve rumen fermentation efficiency in native cattle.

Key words: Yeast, cassava chip, concentrate, rumen fermentation, native cattle

INTRODUCTION

Cassava (*Manihot esculenta*, Crantz) production in tropical areas has a potential use in ruminant livestock nutrition and feeding. Cassava root contains high levels of energy and has been used as a source of readily fermentable energy in ruminant rations (Wanapat, 2003; Kiyothong and Wanapat, 2004; Promkot and Wanapat, 2005). One strategy for using high degradable carbohydrates is to use in combination with readily available NPN sources such as urea. Urea is commonly used as N source when highly soluble carbohydrates are fed and maintained (Wohlt *et al*., 1978). However, efficient utilization of protein and Non-protein Nitrogen (NPN) in ruminants depends upon knowledge of the basic principles underlying ruminal microbial N metabolism (Fernandez *et al*., 1997). Moreover, ruminal pH has great impact on rumen fermentation efficiency (Wanapat, 2003).

In addition, supplementing diets with yeast *(Saccharomyces cerevisiae)* increases milk production of dairy cows and weight gain of growing cattle (Brossard *et al*., 2006). Production responses attributed to yeast are usually related to stimulation of cellulolytic and lactate-utilizing bacteria in the rumen, increased fiber digestion and increased flow of microbial protein from the rumen which may be beneficial for feedlot cattle fed high-grain diets (Guedes *et al*., 2007).

However, the use of yeast fermenting cassava as a replacement for concentrate not yet been investigated. Therefore, the objective of this experiment was to investigate the supplementation of yeast fermenting cassava chip with ruzi grass as a basal roughage on rumen ecology in native cattle.

MATERIALS AND METHODS

Preparation of Yeast Fermented Cassava Chip (YFCC): This technique is based on the method developed by Oboh (2006) and Boonnop *et al*. (2008), which enriching nutritive value of cassava chip with yeast (*Saccharomyces cerevisiae*) fermentation. The method for synthesis of YFCC is as follows:

- Weigh 20 g of yeast in to a flask and add with sugar 20 g, malate 5 g and distill water 100 ml then mixed and incubated at room temperature for 1 h. (A)
- Preparation of medium by weigh 20 g of molasses directly into a warring blender vessel flushed with O_2, add distill water 100 ml and urea 48 g then pour solution and incubated at room temperature for 10 min. (B)
- Adjusting pH media solution by 70% H_2SO_4 between 3.5-.7 and continue mix with incubated for 1 h.
- Remove yeast-malate media solution in a flask from (A) into a medium (B) and continue flush O_2 for 60 h.
- After 60 h, then transfer yeast-malate media solution 50 ml mix with cassava chip 100 g and then covered by plastic bag for a minimum of 72 h.

- Drying of Yeast Fermented Cassava Chip (YFCC) at 30°C for 24 h before feeding to animals.

Animals, diets and experimental design: Ten, one-year old of native cattle weighing at 150±10 kg were randomly divided into two groups according to receive two groups of supplemental feeds by receiving concentrate at 14% CP (T1) and Yeast Fermented Cassava Chip (YFCC). The composition of dietary treatments and ruzi grass used are shown in Table 1.

Cows were housed in individual pens and individually fed concentrate at 1% BW. All cows were fed *ad libitum* of rizi grass with water and a mineral-salt block. Feed intake of concentrate and roughage were measured separately and refusals recorded. The experiment was run for 120 days, the first 15 days for treatment adaptation and for feed intake measurements whist the last 7 days were for sample collections of faeces and rumen fluid. Body weights were measured each 30 days during the sampling period prior to feeding.

Data collection and sampling procedures: Yeast fermented cassava chip, concentrate and ruzi grass were sampled each 30 days and were composted by period prior to analyses. Composites samples were dried at 60°C and ground (1 mm screen using Cyclotech Mill, Tecator, Sweden) and then analyzed for DM, ash and CP content (AOAC, 1985), NDF, ADF and ADL (Goering and Van Soest, 1970).

Rumen fluid and blood samples were collected at 0, 2 and 4 h post-feeding on last period. Approximately 200 ml of rumen fluid was taken from the middle part of the rumen by a stomach tube connected with a vacuum pump at each time at the end of each period. Rumen fluid was immediately measured for pH and temperature using (HANNA instruments HI 8424 microcomputer) after withdrawal. Rumen fluid samples were then filtered through four layers of cheesecloth. Samples were divided into two portions. One portion was used for NH_3-N analyses where 5 ml of H_2SO_4 solution (1M) was added to 50 ml of rumen fluid. The mixture was centrifuged at 16,000 g for 15 min and the supernatant stored at -20°C prior to NH_3-N analysis using the micro Kjeldahl methods (AOAC, 1985). Another portion was fixed with 10% formalin solution in normal saline (Galyean, 1989).

The total count of bacteria, protozoa and fungal zoospores were made using the methods of Galyean (1989) based on the use of a haematocytometer (Boeco). A blood sample (about 10 ml) was drawn from the jugular vein at the same time as rumen fluid sampling, separated by centrifugation at 5,000 g for 10 min and stored at -20°C until analysis of Blood Urea Nitrogen (BUN) according to the method of Crocker (1967).

Statistic analysis: The means of each parameter measured in the digestibility studies and internal parasitic egg counts were analyzed by the analysis of variance procedure of SAS (1998) and means were compared using T-test.

RESULTS AND DISCUSSION

Chemical composition of feeds: The chemical compositions of concentrate diets (T1), yeast fermented cassava chip (YFCC) (T2) and ruzi grass fed in native cattle are shown in Table 1. Crude proteins of concentrate, YFCC and ruzi grass were at 14.2, 29.1 and 8.2%, respectively. Diets containing high levels of cassava chip based diets had a slightly higher Non-structural Carbohydrate (NSC) and lower NDF due to increased level of cassava chip in the diets.

Effect on feed intake and digestibility of nutrients: The effects of supplementation of YFCC as replacement concentrate on feed-intake and digestibility of nutrients in cattle are presented in Table 2. Feed intake were non-significantly different among treatments and was higher in native cattle receiving T2 than T1 (2.7 and 2.5% BW). This result was in agreement with earlier work by (Sommart *et al*., 2000 and Khampa *et al*., 2006) which reported that inclusion of cassava chip in diets resulted in satisfactory animal performance and had no negative effects on animal health in finishing beef cattle and lactating dairy cows.

Characteristics of rumen fermentation and blood metabolism: Rumen ecology parameters were measured for pH and NH_3-N (Table 2). In addition, BUN was determined to investigate their relationships with rumen NH_3-N and protein utilization. Rumen pH at 0, 2 and 4 h post-feeding was changed by dietary treatments, however the values were quite stable at 6.7-6.9, but all treatment means were within the normal range which has been reported as optimal for microbial digestion of fiber and also digestion of protein (6.0-7.0) (Hoover, 1986).

Ruminal NH_3-N and BUN concentrations were altered by YFCC (T2) supplement which containing high cassava-based diets. As NH_3-N is regarded as the most important nitrogen source for microbial protein synthesis in the rumen. In addition, the result obtained was closer to optimal ruminal NH_3-N between at 15-30 mg% (Wanapat and Pimpa, 1999; Chanjula *et al*., 2003, 2004) for increasing microbial protein synthesis, feed digestibility and voluntary feed intake in ruminant fed on low-quality roughages.

Rumen microorganisms populations: Table 3 presents rumen microorganism populations. The populations of fungal zoospores, protozoa and total bacteria direct counts were significantly different and populations of bacteria had higher numbers in native cattle receiving diets YFCC (T2) than T1. In contrast, the present number

Table 1: Chemical composition of concentrate, yeast fermented cassava chip (YFCC) and ruzi grass

Analyzed composition (%)	Concentrate[1]	YFCC	Ruzi grass
DM	91.5	89.1	29.8
OM	90.3	89.5	87.6
CP	14.2	19.2	8.2
TDN	78.3	78.9	57.3
NDF	25.7	17.5	35.6
ADF	14.6	6.1	27.8
ME (Mcal/kg)	3.1	3.3	2.0
Price (US$/kg)	0.28	0.23	0.06

DM = dry matter, CP = crude protein, OM = organic matter, NDF = neutral detergent fiber, ADF = acid detergent fiber, TDN = total digestible of nutrients, ME = metabolizable energy. ([1]Ingredients = concentrate compost of cassava chips 65, fine rice bran 6, brewer's gain 10, palm meal 10, urea 2, molasses 5, sulfur 0.5, salt 0.5 and mineral mix 1%) as dry weight.

Table 2: Effects of supplementation of cattle fed yeast fermented cassava chip (YFCC) as a replacement concentrate on feed intake, blood metabolites (BUN) and ruminal fermentation in native cattle

Item	T1	T2	P-value
DM feed intake (%BW)			
Concentrate	1.0	-	-
YFCC	-	1.0	-
Ruzi grass	1.5	1.7	0.084^{NS}
Total	2.5	2.7	0.147^{NS}
ADG (g/day)	205	259	0.034*
Ruminal fermention			
Temperature (°C)	40.1	39.8	0.423^{NS}
Ruminal pH	6.7	6.9	0.048*
NH_3-N (mg%)	17.6	20.8	0.0341*
BUN (mg%)	9.4	12.1	0.0475*

T1 = Supplementation of concentrate at 14% CP.
T2 = Supplementation of yeast fermented cassava chip (YFCC).
* = $p<0.05$, NS = $p>0.05$

Table 3: Influences of supplementation of yeast fermented cassava chip (YFCC) as a replacement concentrate on rumen microorganisms in cattle

Item	T1	T2	P-value
Total direct counts (cell/ml)			
Bacteria ($x10^{12}$) Protozoa	5.3	9.6	0.0361*
Holotric ($x10^4$)	6.8	4.1	0.0391*
Entodiniomorph ($x10^5$)	8.9	5.2	0.0457*
Fungal zoospores ($x10^6$)	6.3	8.6	0.0414*

T1 = Supplementation of concentrate at 14% CP.
T2 = Supplementation of yeast fermented cassava chip (YFCC).
* = $p<0.05$, NS = $p>0.05$

of protozoa in the rumen was decreased by YFCC supplementation in high cassava-based diets. In the experiment by Guedes *et al.* (2007) reported that yeast are usually related to stimulation of cellulolytic and lactate-utilizing bacteria in the rumen, increased fiber digestion and increased flow of microbial protein from the rumen which may be beneficial for feedlot cattle fed high-grain diets.

Conclusions: Based on this experiment, it could be concluded that supplementation of Yeast Fermented Cassava Chip (YFCC) as a replacement concentrate at 14% CP could improved ruminal fermentation efficiency by increasing populations of bacteria and fungi, but decreased protozoal populations in native cattle.

ACKNOWLEDGEMENTS

The authors wish to express sincere thanks to Rajabhat Mahasarakham University and Tropical Feed Resources Research and Development Center (TROFREC), Khon Kaen University and The National Research Council of Thailand for providing financial support of research and research facilities.

REFERENCES

AOAC., 1985. Official Methods of Analysis. Association of Official Analysis Chemists, DC, USA.

Boonnop, K., M. Wanapat, N. Ngamnit and S. Wanapat, 2008. Enriching Nutritive Value of Cassava Root by Yeast Fermentation. *In:* Proceedings of the graduate school congress x. Held at graduate school khon kaen university, Thailand.

Brossard, L., F. Chaucheyras-Durand, B. Michalet-Doreau and C. Martin, 2006. Dose effect of live yeasts on rumen microbial communities and fermentations during butyric latent acidosis in sheep: newtype of interaction. J. Anim. Sci., 82: 1-11.

Chanjula, P., M. Wanapat, C. Wachirapakorn, S. Uriyapongson and P. Rowlinson, 2003. Ruminal degradability of tropical feeds and their potential use in ruminant diets. Asian-Aust. J. Anim. Sci., 16: 211-216.

Chanjula, P., M. Wanapat, C. Wachirapakorn, S. Uriyapongson and P. Rowlinson, 2004. Effect of synchronizing starch sources and protein (NPN) in the rumen on feed intake, rumen microbial fermentation, nutrient utilization and performance of lactating dairy cows. Asian-Aust. J. Anim. Sci., 17: 1400-1410.

Crocker, C.L., 1967. Rapid determination of urea nitrogen in serum or plasma without deproteinzation. American J. Medical Technol., 33: 361-365.

Fernandez, J.M., T. Sahulu, C. Lu, D. Ivey and M.J. Potchoiba, 1997. Production and metabolic aspects of non-protein nitrogen incorporation in lactation rations of dairy goats. Small Rum. Res., 26: 105-107.

Galyean, M., 1989. Laboratory Procedure in Animal Nutrition Research. Department of Animal and Life Science. New Mexico states University, U.S.A.

Goering, H.K. and P.J. Van Soest, 1970. Forage Fiber Analysis (apparatus, reagent, procedures and some application). Agric. Handbook No. 379, ARS, USDA, Washington, D.C.

Guedes, C.M., D.M. Goncalves, A.M. Rodrigues and A. Dias-da-Silva, 2007. Effects of a *Saccharomyces cerevisiae* yeast on ruminal fermentation and fibre degradation of maize silages in cows. Anim. Feed Sci. Technol., 145: 27-40.

Hoover, W.H., 1986. Chemical factors involved in ruminal fiber digestion. J. Dairy Sci., 69: 2755-2766.

Khampa, S., M. Wanapat, C. Wachirapakorn, N. Nontaso and M. Wattiaux, 2006. Effect of levels of sodium dl-malate supplementation on ruminal fermentation efficiency in concentrates containing high levels of cassava chip in dairy steers. Aisan-Aust. J. Anim. Sci., 19: 368-375.

Kiyothong, K. and M. Wanapat, 2004. Growth hay yield and chemical composition of cassava and Stylo 184 grown under intercropping. Aisan-Aust. J. Anim. Sci., 17: 799-807.

Oboh, G., 2006. Nutrient enrichment of cassava peels using a mixed culture of *Saccharomyces cerevisae* and *Lactobacillus spp* solid media fermentation techniques. Elec. J. Bilo., 9: 46-49.

Promkot, C. and M. Wanapat, 2005. Effect of level of crude protein and use of cottonseed meal in diets containg cassava chips and rice straw for lactating dairy cows. Asian-Aust. J. Anim. Sci., 18: 502-511.

SAS, 1998. SAS/STAT User's Guide. Version 6.12. SAS Inst. Inc., Cary, NC, USA.

Sommart, K., M. Wanapat, D.S. Parker and P. Rowlinson, 2000. Cassava chip as an energy source for lactating dairy cows fed rice straw. Asian-Aust. J. Anim. Sci., 13: 1094-1101.

Wanapat, M., 2003. Manipulation of cassava cultivation and utilization to improve protein to energy biomass for livestock feeding in the tropics. Asian-Aust. J. Anim. Sci., 16: 463-472.

Wanapat, M. and O. Pimpa, 1999. Effect of ruminal NH_3-N levels on ruminal fermentation, purine derivatives, digestibility and rice straw intake in swamp buffaloes. Asian-Aust. J. Anim. Sci., 12: 904-907.

Wohlt, J.E., J.H. Clark and F.S. Blaisdell, 1978. Nutritional value of urea versus preformed protein for ruminants. II. Nitrogen utilization by dairy cows fed corn based diets containing supplemental nitrogen from urea and/or soybean meal. J. Dairy Sci., 61: 916-931.

MEDWELL PUBLISHING Research Journal of Animal Sciences 3 (1): 10-15, 2009
ISSN: 1993-5269

Influences of Supplementation of Yeast-Malate Fermented Cassava Chip as a Replacement Concentrate on Rumen Fermentation Efficiency and Digestibility of Nutrients in Cattle

[1]Sittisak Khampa, [1]Pala Chaowarat, [2]Rungson Singhalert and [3]Metha Wanapat
[1]Faculty of Agricultural Technology, [2]Faculty of Humanities and Social Sciences,
Rajabhat Mahasarakham University, Maha Sarakham, 44000, Thailand
[3]Department of Animal Science, Faculty of Agriculture,
Tropical Feed Resources Research and Development Center,
Khon Kaen University, Khon Kaen, 40002, Thailand

Abstract: Ten, one year old male cattles with initial body weight of 150±10 kg were randomly divided into 2 groups and received concentrate at 14% CP (T_1) and Yeast-Malate Fermented Cassava Chip (YMFCC) (T_2). The cows were offered the treatment concentrate at 1% BW and urea-treated rice straw was fed *ad libitum*. Means were compared using t-test. All animals were kept in individual pens and received free access to water. The results have revealed that replacement of YMFCC on feed intake was non-significantly different, while Average Daily Gain (ADG) and digestibility of nutrients were higher ($p<0.05$) in cattle fed YMFCC (T_2) treatments than received concentrate at 14% CP (T_1) (235 and 203 g day^{-1}). In addition, the ruminal pH, ammonia-nitrogen and blood urea nitrogen concentration were significantly different ($p<0.05$). The concentration of volatile fatty acid was significantly different especially the concentration of propionic acid was slightly higher in cattle receiving T_2 than T_1 (23.9 and 17.8 mol/100 mol). Supplementation of YMFCC (T_2) could improve population of bacteria and fungal zoospore, but decreased populations of *Holotrich* and *Entodiniomorph* protozoa in rumen ($p<0.05$). The results indicate that supplementation of Yeast-Malate Fermented Cassava Chip (YMFCC) as a replacement concentrate at 14% CP could improve rumen fermentation efficiency and digestibility of nutrients in cattle.

Key words: Yeast-Malate, Cassava Chip (YMFCC), concentrate, rumen fermentation, cattle

INTRODUCTION

Cassava (*Manihot esculenta*, Crantz) production in tropical areas has a potential use in ruminant livestock nutrition and feeding. Cassava root contains high levels of energy and has been used as a source of readily fermentable energy in ruminant rations (Wanapat, 2003; Kiyothong and Wanapat, 2004; Promkot and Wanapat, 2005). One strategy for using high degradable carbohydrates is to use in combination with readily available NPN sources such as urea. Urea is commonly used as N source when highly soluble carbohydrates are fed and maintained (Wohlt *et al*., 1978). However, efficient utilization of protein and Non-Protein Nitrogen (NPN) in ruminants depends upon knowledge of the basic principles underlying ruminal microbial N metabolism (Fernandez *et al*., 1997). Moreover, ruminal pH has great impact on rumen fermentation efficiency (Wanapat, 2003). Some strictly anaerobic bacteria use a reductive or reverse citric acid cycle known as the succinate-propionate pathway to synthesize succinate and (or) propionate. Both malate and fumalate are key intermediates in the succinate propionate pathway and *S. ruminantium* uses this pathway (Gottschalk, 1986). The fact dicarboxylic acids, especially malate and fumalate, stimulate lactate utilization is consistent with the presence of this pathway in this ruminal anaerobe (Callaway and Martin, 1996). Previous studies by Sanson and Stallcup (1984) reported that supplementation of malate in ruminant diets has been shown to increase nitrogen retention in sheep and steers and to improve average daily gain and feed efficiency in bull calves. In addition, supplementing diets with yeast (*Saccharomyces cerevisiae*) increases milk production of dairy cows and weight gain of growing cattle (Brossard *et al*., 2006). Production responses attributed to yeast are usually related to stimulation of cellulolytic and lactate-utilizing bacteria in the rumen, increased fiber digestion and increased flow of microbial protein from the rumen which may be beneficial for feedlot cattle fed high-grain diets (Guedes, 2007).

Corresponding Author: Sittisak Khampa, Faculty of Agricultural Technology, Rajabhat Mahasarakham University, Maha Sarakham, 44000, Thailand

However, the use of yeast-malate fermenting cassava as a replacement for concentrate not yet been investigated. Therefore, the objective of this experiment was to investigate the supplementation of yeast-malate fermenting cassava with rice straw as a basal roughage on rumen fermentation efficiency and growth in cattle.

MATERIALS AND METHODS

Preparation of Yeast-Malate Fermented Cassava Chip (YMFCC): This technique is based on the method developed by Oboh (2006) and Boonnop *et al.* (2008), which enriching nutritive value of cassava chip with yeast (*Saccharomyces cerevisiae*) fermentation. The method for synthesis of YMFCC is as follows:

- Weigh 20 g of yeast into a flask and add with sugar 20 g, malate 5 g and distill water 100 mL then mixed and incubated at room temperature for 1 h (A)
- Preparation of medium by weigh 20 g of molasses directly into a warring blender vessel flushed with O_2, add distill water 100 mL and urea 48 g then pour solution and incubated at room temperature for 10 min (B)
- Adjusting pH media solution by 70% H_2SO_4 between 3.5-0.7 and continue mix with incubated for 1 h
- Remove yeast-malate media solution in a flask from (A) into a medium (B) and continue flush O_2 for 60 h
- After 60 h, then transfer yeast-malate media solution 50 mL mix with cassava chip 100 g and then covered by plastic bag for a minimum of 72 h
- Drying of Yeast-Malate Fermented Cassava Chip (YMFCC) at 30°C for 24 h before feeding to animals

Animals, diets and experimental design: Ten, one year old of male cattles weighing at 150±10 kg were randomly divided into 2 groups according to receive 2 groups of supplemental feeds by receiving concentrate at 14% CP (T_1) and Yeast-Malate Fermented Cassava Chip (YMFCC). The composition of dietary treatments and Urea-treated Rice Straw (UTS) used are shown in Table 1 and 2.

Cows were housed in individual pens and individually fed concentrate at 1% BW. All cows were fed *ad libitum* of UTS with water and a mineral-salt block. Feed intake of concentrate and roughage were measured separately and refusals recorded. The experiment was run for 120 days, the first 15 days for treatment adaptation and for feed intake measurements whist the last 7 days were for sample collections of faeces, urine and rumen fluid. Body weights were measured each 30 days during the sampling period prior to feeding.

Table 1: Ingredients of concentrate used in the experiment (DM% basis)

Ingredients (DM%)	Concentrate
Cassava chip	65.0
Fine rice bran	6.0
Brewer's grain	10.0
Palm meal	10.0
Urea	2.0
Molasses	5.0
Sulfur	0.5
Salt	0.5
Mineral mix	1.0

Table 2: Chemical composition of concentrate, Yeast-Malate Fermented Cassava Chip (YMFCC) and Urea-Treated rice Straw (UTS)

Analyzed composition (%)	Concentrate	YMFCC	UTS
DM	91.50	89.10	55.80
OM	90.30	89.50	88.90
CP	14.20	29.10	7.90
TDN[1]	78.30	78.90	55.10
NDF	25.70	17.50	73.20
ADF	14.60	6.10	52.30
ME (Mcal kg^{-1})	3.10	3.30	1.90
Price (US$ kg^{-1})	0.28	0.23	0.05

[1]TDN = dig CP + Dig CF + dig EE × 2.25 + dig NFE

UTS was prepared by using 5% (w w^{-1}) urea mixed with 100 kg of water in 100 kg of Rice Straw (RS) batches (50:50, water to straw) and poured over a stack of straw and then covered with a plastic sheet for a minimum of 10 days before feeding to animals (Wanapat, 1990).

Data collection and sampling procedures: UTS, YMFCC and concentrate diets were sampled each 30 days and were composted by period prior to analyses. Feed, fecal and urine samples were collected by rectal sampling whist urine samples were collected by spot sampling during the last seven days of each period. Composites samples were dried at 60°C and ground (1 mm screen using Cyclotech Mill, Tecator, Sweden) and then analyzed for DM, ether extract, ash and CP content (AOAC, 1990), NDF, ADF and ADL (Van Soest *et al.*, 1991) and AIA. AIA was used to estimate digestibility of nutrients (Van Keulen and Young, 1977).

Rumen fluid and blood samples were collected at 0, 2 and 4 h post-feeding on last period. Approximately 200 mL of rumen fluid was taken from the middle part of the rumen by a stomach tube connected with a vacuum pump at each time at the end of each period. Rumen fluid was immediately measured for pH and temperature using (HANNA instruments HI 8424 microcomputer) after withdrawal. Rumen fluid samples were then filtered through 4 layers of cheesecloth. Samples were divided into 2 portions. One portion was used for NH_3-N analyses where 5 mL of H_2SO_4 solution (1 M) was added to 50 mL of rumen fluid. The mixture was centrifuged at 16,000 g for 15 min and the supernatant stored at -20°C prior to NH_3-N analysis using the micro Kjeldahl methods (AOAC, 1990) and Volatile Fatty Acids (VFAs) analyses using a HPLC

according to Zinn and Owens (1986). Another portion was fixed with10% formalin solution in normal saline (Galyean, 1989).

The total count of bacteria, protozoa and fungal zoospores were made using the methods of Galyean (1989) based on the use of a haematocytometer (Boeco). A blood sample (about 10 mL) was drawn from the jugular vein at the same time as rumen fluid sampling, separated by centrifugation at 5,000 g for 10 min and stored at -20°C until analysis of Blood Urea Nitrogen (BUN) according to the method of Crocker (1967).

Statistic analysis: The means of each parameter measured in the digestibility studies and internal parasitic egg counts were analyzed by the analysis of variance procedure of SAS (1998) and means were compared using t-test.

RESULTS AND DISCUSSION

Chemical composition of feeds: The chemical compositions of concentrate diets (T_1), Yeast-Malate Fermented Cassava Chip (YMFCC) (T_2) and Urea-Treated rice Straw (UTS) fed in cattle are shown in Table 2. Crude proteins of concentrate, YMFCC and UTS were at 14.2, 29.1 and 7.9%, respectively. Diets containing high levels of cassava chip based diets had a slightly higher Non-Structural Carbohydrate (NSC) and lower NDF due to increased level of cassava chip in the diets. Furthermore, the chemical composition of UTS is presented in Table 2. Similar values for UTS have been similar to those reported by Wanapat (2000).

Effect on feed intake and digestibility of nutrients: The effects of supplementation of YMFCC as replacement concentrate on feed-intake and digestibility of nutrients in cattle are presented in Table 3. Feed intake were non-significantly different among treatments and was higher in cattles receiving T_2 than T_1 (2.6 and 2.5% BW). This result was in agreement with earlier work by (Sommart *et al.*, 2000; Khampa *et al.*, 2006), which reported that inclusion of cassava chip in diets resulted in satisfactory animal performance and had no negative effects on animal health in finishing beef cattle and lactating dairy cows.

Apparent digestibility of DM, OM, CP, NDF and ADF were non-significant different ($p<0.05$) for all diets, however digestible of nutrient intake tended te be higher in cattle fed YMFCC (T_2) than T_1. The slightly lower NDF digestibility of the cassava-based diets may have contributed to higher degradation in substantial decrease in fiber digestibility as reported by Hoover (1986).

Table 3: Apparent digestibility and feed-intake of cattle fed Yeast-Malate Fermented Cassava Chip (YMFCC) as a replacement concentrate

Item	T_1	T_2	p-value
DM Intake (BW%)			
Concentrate	1.0	-	-
YMFCC	-	1.0	-
Rice straw	1.5	1.6	0.7732^{ns}
Total	2.5	2.6	0.6841^{ns}
Apparent digestibility (%)			
DM	65.7	67.1	0.521^{ns}
OM	68.5	71.2	0.987^{ns}
CP	74.3	76.3	0.536^{ns}
NDF	62.4	64.9	0.742^{ns}
ADF	47.2	49.1	0.856^{ns}
ADG (g day^{-1})	203	235	0.0278*
Cost production (US$ kg^{-1} BW)	2.94	2.4	0.0351*

T_1 = Supplementation of concentrate at 14% CP; T_2 = Supplementation of Yeast-Malate Fermented Cassava Chip (YMFCC); *p<0.05; ns: p>0.05

Furthermore, in the experiment by Erdman (1998) reported that the sources of starch influence the rate of NDF digestion differently at pH 6.8 than 5.5. In addition, when ruminal pH was reduced below 6.3 in dairy cows, ADF digestion could be decreased at 3.6% unit per 0.1 pH and may result in depressed feed-intake.

Characteristics of ruminal fermentation and blood metabolism: Rumen ecology parameters were measured for pH, NH_3-N and VFA (Table 4). In addition, BUN was determined to investigate their relationships with rumen NH_3-N and protein utilization. Rumen pH at 0, 2 and 4 h post-feeding was changed by dietary treatments, however the values were quite stable at 6.6-6.9, but all treatment means were within the normal range which has been reported as optimal for microbial digestion of fiber and also digestion of protein (6.0-7.0) (Hoover, 1986).

Ruminal NH_3-N and BUN concentrations were altered by YMFCC (T_2) supplement, which containing high cassava-based diets. As NH_3-N is regarded as the most important nitrogen source for microbial protein synthesis in the rumen. In addition, the result obtained was closer to optimal ruminal NH_3-N between at 15-30 mg% (Wanapat and Pimpa, 1999; Chanjula *et al.*, 2003, 2004) for increasing microbial protein synthesis, feed digestibility and voluntary feed intake in ruminant fed on low-quality roughages.

The influence of supplementation of Yeast-Malate Fermented Cassava Chip (YMFCC) as a replacement concentrate on production of total VFA, acetic acid proportion, propionic acid proportion, butyric acid proportion and acetic to propionic ratio are shown in Table 4. Mean total VFAs and propionate concentrations in the rumen were significantly different by increased with receiving YMFCC (T_2) than T_1 (117.6 and 102.4 mM). However, it was found that total VFA concentration in all diets ranged from 70-130 mM, the range suggested by France and Siddons (1993). Especially, the acetate to

Table 4: Influences of supplementation of Yeast-Malate Fermented Cassava Chip (YMFCC) as a replacement concentrate on rumen fermentation and blood metabolites in cattle

Item	T_1	T_2	P-value
Ruminal pH	6.6	6.9	0.0372*
NH_3-N (mg%)	17.2	21.4	0.0432*
BUN (mg%)	8.6	13.4	0.0457*
Total VFA (mM L^{-1})	102.4	117.6	0.0351*
Molar proportion of VFA (mol/100 mol)			
Acetate (C2)	72.4	66.8	0.0481*
Propionate (C3)	17.8	23.9	0.0531*
Butyrate (C4)	9.8	9.3	0.0842^{ns}
C2:C3 ratio	4.1	2.7	0.0412*
C2+C4:C3 ratio	4.6	3.1	0.0429*

T_1 = Supplementation of concentrate at 14% CP; T_2 = Supplementation of Yeast-malate Fermented Cassava Chip (YMFCC); *p<0.05; ns: p>0.05

Table 5: Influences of supplementation of Yeast-Malate Fermented Cassava Chip (YMFCC) as a replacement concentrate on rumen microorganisms in cattle

Item	T_1	T_2	p-value
Total direct counts (cell mL^{-1})			
Bacteria ($\times 10^{11}$)	6.8	8.4	0.0452*
Protozoa Holotric ($\times 10^{5}$)	6.5	4.6	0.0463*
Entodiniomorph ($\times 10^{5}$)	5.1	2.7	0.0374*
Fungal zoospores ($\times 10^{6}$)	4.9	6.8	0.0472*

T_1 = Supplementation of concentrate at 14% CP; T_2 = Supplementation of Yeast-Malate Fermented Cassava Chip (YMFCC); *p<0.05

propionate ratio was decreased by receiving YMFCC (T_2) than T_1 (2.7 and 4.1) but the supplementation of YMFCC (T_2) increased the daily output of propionate without decreasing the production of acetate (23.9 and 17.8 mol/100 mol) and it was in agreement with the results reported by other authors (Callaway and Martin, 1996; Khampa *et al.*, 2006).

Rumen microorganisms populations: Table 5 presents rumen microorganism populations. The populations of fungal zoospores, protozoa and total bacteria direct counts were significantly different and populations of bacteria had higher numbers in cattle receiving diets YMFCC (T_2) than T_1. In contrast, the present number of protozoa in the rumen was decreased by YMFCC supplementation in high cassava-based diets. In the experiment by Newbold *et al.* (1996) has shown that feeding 100 mg of malate day^{-1} in sheep caused an increase in the number of total bacteria and tended to increase the population of cellulolytic bacteria. In agreement with these observations, Lopez *et al.* (1999) reported that fumalate (another intermediate in the succinate to propionate pathway) increased the number of cellulolytic bacteria almost three-fold during fermentation in the RUSITEC system. In addition Guedes (2007) reported that yeast are usually related to stimulation of cellulolytic and lactate-utilizing bacteria in the rumen, increased fiber digestion and increased flow of microbial protein from the rumen which may be beneficial for feedlot cattle fed high-grain diets. As cassava chip can be readily degraded in the rumen and ruminal pH was decreased, malate could stimulate lactate utilization by *S. ruminantium* and could improve pH in the rumen. It is possible that supplementation of malate with yeast may play an important role in increasing bacterial populations. Moreover, Martin *et al.* (1999) reported that increasing dietary concentrations of malate might help to reduce problems associated with ruminal.

CONCLUSION

Based on this experiment, it could be concluded that supplementation of Yeast-Malate Fermented Cassava Chip (YMFCC) as a replacement concentrate at 14% CP could improved ruminal fermentation efficiency, digestibility of nutrients and increasing propionate production, but decreased acetate to propionate ratio. In addition, supplementation of YMFCC increase populations of bacteria, but decreased protozoal populations in cattle.

ACKNOWLEDGEMENT

The authors wish to express sincere thanks to Rajabhat Mahasarakham University and Tropical Feed Resources Research and Development Center (TROFREC), Khon Kaen University and The National Research Council of Thailand for providing financial support of research and research facilities.

REFERENCES

AOAC, 1990. Official Methods of Analysis. Association of Official Analysis Chemists. 15th Edn. Inc. Arlington, Virginia. ISBN: 0-935584-42-0.

Boonnop, K., M. Wanapat, N. Ngamnit and S. Wanapat, 2008. Enriching Nutritive Value of Cassava Root by Yeast Fermentation. Proceedings of the graduate school congress X. Held at graduate school Khon Kaen University, Thailand, pp: 97-102.

Brossard, L., F. Chaucheyras-Durand, B. Michalet-Doreau and C. Martin, 2006. Dose effect of live yeasts on rumen microbial communities and fermentations during butyric latent acidosis in sheep: Newtype of interaction. J. Anim. Sci., 82: 1-11. DOI: 10.1017/ASC200693.

Callaway, T.R. and S.A. Martin, 1996. Effects of organic acid and monensin treatment on *in vitro* mixed ruminal microorganism fermentation of cracked corn. J. Anim. Sci., 74: 1982-1989. http://jas.fass.org/cgi/reprint/74/8/1982.pdf.

Chanjula, P., M. Wanapat, C. Wachirapakorn, S. Uriyapongson and P. Rowlinson, 2003. Ruminal degradability of tropical feeds and their potential use in ruminant diets. Asian-Aust. J. Anim. Sci., 16: 211-216. http://cat.inist.fr/?aModele=afficheN&cpsidt=15029234.

Chanjula, P., M. Wanapat, C. Wachirapakorn, S. Uriyapongson and P. Rowlinson, 2004. Effect of synchronizing starch sources and protein (NPN) in the rumen on feed intake, rumen microbial fermentation, nutrient utilization and performance of lactating dairy cows. Asian-Aust. J. Anim. Sci., 17: 1400-1410. www.pjbs.org/pjnonline/fin542.pdf.

Crocker, C.L., 1967. Rapid determination of urea nitrogen in serum or plasma without deproteinzation. Am. J. Med. Technol., 33: 361-365. www.pjbs.org/pjnonline/fin470.pdf.

Erdman, R.A., 1998. Dietary buffering requirements of the lactating dairy cows. A review. J. Dairy Sci., 71: 3246-3266. http://jds.fass.org/cgi/reprint/71/12/3246.pdf.

Fernandez, J.M., T. Sahulu, C. Lu, D. Ivey and M.J. Potchoiba, 1997. Production and metabolic aspects of non-protein nitrogen incorporation in lactation rations of dairy goats. Small Rum. Res., 26: 105-107. DOI: 10.1016/S0921-4488(96)00984-4.

France, J. and R.C. Siddons, 1993. Volatile fatty acid production. Quantilitive Aspects Ruminant Digestion and Metabolisim. In: Forbes J.M. and J. France (Eds.) C.A.B. International, Willingford, UK., pp: 196-203. ISBN: 9780851998145.

Galyean, M., 1989. Laboratory Procedure in Animal Nutrition Research New Mexico states University, USA., pp: 162-167.

Gottschalk, G. 1986. Bateria Metabolism. 2nd Edn. Sparinger-Verlag. New York. ISBN-13: 9780387903088.

Guedes, C.M., D.M. Goncalves, A.M. Rodrigues and A. Dias-da-Silva, 2007. Effects of a *Saccharomyces cerevisiae* yeast on ruminal fermentation and fibre degradation of maize silages in cows. Anim. Feed Sci. Technol., 145: 27-40. http://linkinghub.elsevier.com/retrieve/pii/S0377840107002866.

Hoover, W.H., 1986. Chemical factors involved in ruminal fiber digestion. J. Dairy Sci., 69: 2755-2766. PMID: 3027148. http://jds.fass.org/cgi/reprint/69/10/2755.pdf.

Khampa, S., M. Wanapat, C. Wachirapakorn, N. Nontaso and M. Wattiaux, 2006. Effect of levels of sodium dl-malate supplementation on ruminal fermentation efficiency in concentrates containing high levels of cassava chip in dairy steers. Aisan-Aust. J. Anim. Sci., 19: 368-375. http://cat.inist.fr/?aModele=afficheN&cpsidt=17613083.

Kiyothong, K. and M. Wanapat, 2004. Growth, hay yield and chemical composition of cassava and Stylo 184 grown under intercropping. Aisan-Aust. J. Anim. Sci., 17: 799-807. www.pjbs.org/pjnonline/fin542.pdf.

Lopez, S., C. Newbold and R.J. Wallace, 1999. Influence of sodium fumarate addition on rumen fermentation *in vitro*. Br. J. Nutr. 81: 59-64. http://journals.cambridge.org/download.php?file=%2FBJN%2FBJN81_01%2FS0007114599000015Xa.pdf&code=fbcd8c2476b2974513924f0d065b8fde.

Martin, S.A., M.N. Streeter, D.J. Nisbet, G.M. Hill and E.E. Williams, 1999. Effect of DL-malate on ruminal metabolism and performance of cattle fed a high concentrate diets. J. Anim. Sci., 77: 1008-1015. http://jas.fass.org/cgi/reprint/77/4/1008.pdf.

Newbold, C.J., R.J. Wallace and F.M. McIntosh, 1996. Mode of action of the yeast *Saccharomyces cerevisiae* as a feed additive for ruminants. Br. J. Nutr., 76: 249-261. http://journals.cambridge.org/download.php?file=%2FBJN%2FBJN76_02%2FS0007114596001328a.pdf&code=3be260eaa21a4ec41d84a553e6b92ef1.

Oboh, G., 2006. Nutrient enrichment of cassava peels using a mixed culture of *Saccharomyces cerevisae* and *Lactobacillus* sp. solid media fermentation techniques. Elec. J. Biol., 9: 46-49. http://www.bioline.org.br/abstract?id=ej06006&lang=es.

Promkot, C. and M. Wanapat, 2005. Effect of level of crude protein and use of cottonseed meal in diets containg cassava chips and rice straw for lactating dairy cows. Asian-Aust. J. Anim. Sci., 18: 502-511. http://cat.inist.fr/?aModele=afficheN&cpsidt=16601064.

Sanson, D.W. and O.T. Stallcup, 1984. Growth response and serum constituents of Holstein bulls fed malic acid. Nutr. Rep. Int., 30: 1261-1267. http://pt.wkhealth.com/pt/re/bjon/abstract.00002375-200302000-00003.htm;jsessionid=J4ZT41z9T5dstP2yWjSBWkbgRjGhsJLm6g6NWv5cbGTVMcQVLnyG!-185407979!181195628!8091!-1.

SAS, 1998. SAS/STAT User's Guide. Version 6.12. SAS Inst. Inc., Cary, NC. ISBN: 1-55544-3561.

Sommart, K., M. Wanapat, D.S. Parker and P. Rowlinson, 2000. Cassava chip as an energy source for lactating dairy cows fed rice straw. Asian-Aust. J. Anim. Sci., 13: 1094-1101. http://grande.nal.usda.gov/ibids/index.php?mode2=detail&origin=ibids_references&therow=455517.

Van Keulen, J. and B.A. Young, 1977. Evaluation of acid insoluble ash as a neutral marker in ruminant digestibility studies. J. Anim. Sci., 44: 282-287. http://jas.fass.org/cgi/reprint/44/2/282.pdf.

Van Soest, P.J., J.B. Robertson and B.A. Lewis, 1991. Methods for Dietary Fiber, Neutral Detergent Fiber and Nonstarch Polysaccharides in Relation to Animal Nutrition. J. Dairy Sci., 743583-3597. http://jds.fass.org/cgi/reprint/74/10/3583.pdf.

Wanapat, M., 1990. Nutritional Aspects of Ruminant Production in Southeast Asia with Special Reference to Thailand. Funny Press, Ltd., Bangkok, Thailand, pp: 125-136.

Wanapat, M., 2000. Rumen manipulation to increase the efficient use of local feed resources and productivity of ruminants in the tropics. Asian-Aust. J. Anim. Sci., 13: 59-67. www.pjbs.org/pjnonline/fin589.pdf.

Wanapat, M., 2003. Manipulation of cassava cultivation and utilization to improve protein to energy biomass for livestock feeding in the tropics. Asian-Aust. J. Anim. Sci., 16: 463-472. http://cat.inist.fr/?aModele=afficheN&cpsidt=15035124.

Wanapat, M. and O. Pimpa, 1999. Effect of ruminal NH_3-N levels on ruminal fermentation, purine derivatives, digestibility and rice straw intake in swamp buffaloes. Asian-Aust. J. Anim. Sci., 12: 904-907. http://grande.nal.usda.gov/ibids/index.php?mode2=detail&origin=ibids_references&therow=454349.

Wohlt, J.E., J.H. Clark and F.S. Blaisdell, 1978. Nutritional value of urea versus preformed protein for ruminants. II. Nitrogen utilization by dairy cows fed corn based diets containing supplemental nitrogen from urea and/or soybean meal. J. Dairy Sci., 61: 902-905. http://jds.fass.org/cgi/reprint/61/7/902.pdf.

Zinn, A.R. and F.N. Owen, 1986. A rapid procedure for purine measurement and its use for estimating net ruminal protein synthesis. Can. J. Anim. Sci., 66: 157-163.http://www-naweb.iaea.org/nafa/aph/public/aph-tecdoc-945.pdf.